

## **Technical Information**

### **Phenol Red Adonitol Broth**

### Product Code: DM 2200

Application: - Phenol Red Adonitol Broth is used for detection of adonitol fermenting bacteria.

### Composition\*\*

| Ingredients   | Gms / Litre |
|---|-------------|
| Proteose peptone  | 10.000      |
| Beef extract  | 1.000       |
| Sodium chloride   | 5.000       |
| Adonitol  | 5.000       |
| Phenol red  | 0.018       |
| Final pH ( at 25°C)   | 7.4±0.2     |
| **Formula adjusted, standardized to suit performance parameters |             |

# **Principle & Interpretation**

Phenol Red Broth Medium devised by Vera <sup>(2)</sup> is recommended to determine the fermentation behavior of different carbohydrates for the differentiation of microorganisms <sup>(3-5)</sup>. Phenol Red Broth Medium with various carbohydrates serves as a differential medium by helping in differentiation of various species and genera by their ability to ferment the specific carbohydrate, with the production of acid or acid and gas <sup>(6)</sup>. Phenol Red Adonitol Broth is used to study adonitol fermentation in various bacteria.

Proteose peptone and beef extract serve as sources for carbon and nitrogen. Sodium chloride is the osmotic stabilizer. Phenol red is the pH indicator, which turns yellow at acidic pH i.e. on fermentation of adonitol. Gas formation is seen in Durham's tubes. All of the Enterobacteriaceae grow well in this medium. In addition to producing a pH colour shift, the production of mixed acids, notably butyric acids, often results in a pungent, foul odour from the culture medium (1).

## Methodology

Suspend 21 grams of powder media in 1000 ml distilled water and mix well. Shake well & heat if necessary to ensure complete solution. Distribute in fermentation tubes (tubes containing inverted Durham's tubes). Sterilize by autoclaving at 15 lbs pressure (121°C) for 15 minutes.

## **Quality Control**

#### Physical Appearance

Light yellow to pink coloured homogeneous free flowing powder

#### Colour and Clarity of prepared medium

Red coloured clear solution without any precipitate

#### Reaction

Reaction of 2.1% w/v aqueous solution at 25°C.  $7.4\pm0.2$  pH :  $7.4\pm0.2$ 

#### pH Range

7.20-7.60

#### Cultural Response/Characteristics

DM 2200: Cultural characteristics observed after an incubation at 35-37°C for 18-24 hours.

| Organism                          | Inoculum<br>(CFU) | Growth    | Acid<br>production                  | Gas               |
|-----------------------------------|-------------------|-----------|-------------------------------------|-------------------|
| Citrobacter freundii<br>ATCC 8090 | 50-100            | Luxuriant | Negative reaction, no colour change | Negative reaction |





| Escherichia coli<br>ATCC 25922       | 50-100 | luxuriant | Negative reaction, no colour change | Negative reaction |
|--------------------------------------|--------|-----------|-------------------------------------|-------------------|
| Enterobacter aerogenes<br>ATCC 13048 | 50-100 | Luxuriant | Positive reaction, yellow colour    | Positive reaction |
| Klebsiella pneumoniae<br>ATCC 13883  | 50-100 | Luxuriant | Positive reaction, yellow colour    | Positive reaction |
| Proteus vulgaris<br>ATCC 13315       | 50-100 | Luxuriant | Negative reaction, no colour change | Negative reaction |
| Serratia marcescens<br>ATCC 8100     | 50-100 | Luxuriant | Negative reaction, no colour change | Negative reaction |
| Salmonella Typhi<br>ATCC 6539        | 50-100 | Luxuriant | Negative reaction, no colour change | Negative reaction |
| Salmonella Typhimurium<br>ATCC 14028 | 50-100 | Luxuriant | Negative reaction, no colour change | Negative reaction |
| Shigella flexneri<br>ATCC 12022      | 50-100 | Luxuriant | Negative reaction, no colour change | Negative reaction |

## Storage and Shelf Life

**Dried Media:** Store below 30°C in tightly closed container and use before expiry date as mentioned on the label. **Prepared Media:** 2-8<sup>0</sup> in sealable plastic bags for 2-5 days.

## **Further Reading**

- 1. Koneman E. W., Allen S. D., Janda W.M., Schreckenberger P.C., Winn W.C. Jr., 1992, Colour Atlas and Textbook of Diagnostic Microbiology, 4th Ed., J. B. Lippinccott Company
- 2. Vera H. D., 1950, Am. J. Public Health, 40, 1267
- 3. MacFaddin J. F., 1985, Media for Isolation-Cultivation-Identification -Maintenanceof Medical Bacteria, Vol. I, Williams and Wilkins, Baltimore.
- 4. Finegold S. M. and Baron E. J., 1986, Bailey and Scotts Diagnostic Microbiology, 7th Ed., The C.V. Mosby Co., St. Louis.
- 5. Ewing W. H., 1986, Edwards and Ewings Identification of Enterobacteriaceae, 4th ed., Elsevier Science Publishing Co., Inc., New York.
- 6. MacFaddin J. F., 2000, Biochemical tests for Identification of Medical Bacteria, 3rd edi., Lippincott, Williams and Wilkins, Baltimore.

### Disclaimer :

- User must ensure suitability of the product(s) in their application prior to use.
- The product conform solely to the technical information provided in this booklet and to the best of knowledge research and development work carried at **CDH** is true and accurate
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